## Sensing and signalling intercalary growth in *Epichloë*endophytes: A transcriptomics approach to mechanosensing.

A. Ozturk<sup>1</sup>, S. Ariyawansa<sup>1</sup>, R. E. Bradshaw<sup>2</sup>, P. Maclean<sup>1</sup>, J. Koolaard<sup>1</sup>, P. Dupont<sup>2</sup>, C. J. Eaton<sup>2</sup>, N. D. Read<sup>3</sup>, N. A. Gow<sup>4</sup>, C. R. Voisey<sup>1</sup>

AgResearch, Grasslands Research Centre, Private Bag 11-008, Palmerston North 4442, New Zealand; 2) Bio-Protection Research Centre, Institute of Fundamental Sciences, Massey University, Palmerston North 4474, New Zealand; 3) Manchester Fungal Infection Group, Division of Infection, Immunity and Respiratory Medicine, University of Manchester, Manchester M13 9NT, United Kingdom; 4) The Aberdeen Fungal Group, School of Medicine, Medical Science and Nutrition, Institute of Medical Sciences, University of Aberdeen, Aberdeen AB252ZD, UK.

Although fungal hyphae normally extend at apical tips, exceptions to polar growth characterise the ecology of many important species. We are investigating the physiology of intercalary growth in Epichloëfestucae, a seed-transmitted symbiont that colonises the aerial parts of grasses. Developing leaves are initially invaded by hyphaltips, however continued hyphalcolonisation occurs through expansion of intercalary compartments. We recently tested the hypothesis that intercalary growth is stimulated by mechanical forces imposed on hyphae through their attachment to expanding host cells. E. festucae hyphae were grown in vitro on silicon membranes and stretching forces were shown to induce intercalary compartment extension and division, as observed in developing leaves. The aim of this study was to identify the cellular responses that induce intercalary hyphal growth after stretch. E. festucae cultures were grown on silicon membranes fitted to a custom-designed stretching frame, stretched by 8%, and then harvested either 5 min or 3 h later. The transcriptomes of stretched hyphae were compared against controls grown on the same devices with no stretch applied. Over 100 genes were differentially expressed in cultures harvested 5 min after stretch, whereas after 3 h over 400 genes were differentially regulated. The results suggest that reprogramming of primary metabolism and plasma membrane organisation occurs almost immediately in response to mechanical stress, and mobilisation of cell wall enzymes and secondary metabolism is observed in the cultures harvested later. These findings will assist in determining the role of mechanical stress in regulating *E. festucae* physiology and growth in plants.